RESEARCH ARTICLE

Hyperprogesteronemia in Response to Vitex fischeri Consumption in Wild Chimpanzees (Pan troglodytes schweinfurthii)

MELISSA EMERY THOMPSON^{1,2*}, MICHAEL L. WILSON^{3,4}, GRACE GOBBO⁵, MARTIN N. MULLER¹, AND ANNE E. PUSEY⁴

^{AND} ANNE E. FOSE1 ¹Department of Anthropology, University of New Mexico, Albuquerque, New Mexico ²Department of Anthropology, Harvard University, Cambridge, Massachusetts ³Department of Anthropology, University of Minnesota, Minneapolis, Minnesota ⁴Department of Ecology, Evolution and Behavior, University of Minnesota, Minneapolis, Minnesota ⁵Gombe Stream Research Centre, The Jane Goodall Institute, Kigoma, Tanzania

Chimpanzees in Gombe National Park consume fruits of *Vitex fischeri* during a short annual fruiting season. This fruit species is a member of a genus widely studied for phytoestrogen composition and varied physiological effects. One particularly well-studied species, V. agnus-castus, is noted for its documented effects on female reproductive function, evidenced in increased progesterone levels and consequent regulation of luteal function. We examined reproductive hormone levels in both male and female chimpanzees during a 6-week period of intense V. fischeri consumption. V. fischeri consumption was associated with an abrupt and dramatic increase in urinary progesterone levels of female chimpanzees to levels far exceeding the normal range of variation. Female estrogen levels were not significantly impacted, nor were male testosterone levels. These are some of the first data indicating that phytochemicals in the natural diet of a primate can have significant impacts on the endocrine system, though the fluctuating nature of chimpanzee diet and reproductive function does not allow us to determine whether the effects observed during this short period had a broader positive or negative impact on female fertility. Given the widespread use of various Vitex species by African primates and the as-yet-undescribed phytochemical properties of these species, we predict that our observations may be indicative of a broader phenomenon. Am. J. Primatol. 70:1064–1071, 2008. © 2008 Wiley-Liss, Inc.

Key words: diet; phytoestrogens; reproduction; progesterone; chimpanzees

INTRODUCTION

There is a robust literature on the potential for phytoestrogens and other chemical constituents of natural plant foods to influence the reproductive physiology of humans and other animals. Phytoestrogens, chemicals having structural and/or functional similarities to endogenous estrogens, can influence the reproductive system via several pathways, including steroid receptor densities and affinities, cell signaling pathways, cell proliferation, and steroid biosynthesis [Whitten & Naftolin, 1998; Whitten & Patisaul, 2001]. Specific reproductive effects have been demonstrated from administered diets rich in phytoestrogens, though the nature of these effects vary and may be positive or negative depending on the specific compounds administered, doses, and the timing and the duration of administration [review: Whitten & Patisaul, 2001]. Naturally phytoestrogen-rich diets have a variety of inhibitory effects on reproduction in herbivores [Adams, 1995; Hughes, 1988] and quail [Leopold et al., 1976]. Evidence for phytochemical influences on the reproductive physiology of primates is primarily limited to laboratory studies [Foth & Cline, 1998; Harrison et al., 1999; Hopkins et al., 1977; Trisomboon et al., 2004; Wood et al., 2006]. However, Whitten [1983] noted a close correlation of seasonal mating behavior in wild vervet monkeys (Cercopithecus aethiops) with the consumption of estrogenic Acacia elatior flowers [Garey et al., 1992], and other researchers have speculated that the conspicuous consumption of particular foods rich in phytochemicals may alter the reproductive function of their study animals [Strier, 1993; Wallis, 1997]. Recently-published research documents elevated progesterone in response to consumption of Vitex doniana fruits by baboons in Gashaka-Gumti National Park, Nigeria

Received 17 April 2008; revision accepted 27 June 2008

DOI 10.1002/ajp.20600

Contract grant sponsors: Jane Goodall Institute; University of Minnesota; US National Science Foundation; Harris Steel Group; Harvard University; Wenner-Gren Foundation; L. S. B. Leakev Foundation.

^{*}Correspondence to: Melissa Emery Thompson, Department of Anthropology, MSC01-1040, University of New Mexico, Albuquerque, NM 87131-0001. E-mail: memery@unm.edu

Published online 21 July 2008 in Wiley InterScience (www. interscience.wiley.com).

(Higham et al., 2007). In this study, we provide strong evidence for elevation of progesterone levels in female chimpanzees as the direct result of the consumption of the fruit *Vitex fischeri*.

Researchers have described a wide range of biological effects of phytoestrogens and other compounds contained in several members of the plant genus Vitex [e.g., V. negundo: Bhargava, 1989; V. doniana: Eckman & Hines, 1993; V. polygama: Goncalves et al., 2001; V. trifolia: Li et al., 2005; V. rotundifolia: Okuyama et al., 1998; V. altissima: Sridhar et al., 2005; V. peduncularis: Suksamrarn et al., 2002]. These plants and their constituents have variously been cited for their effectiveness as antiinflammatory [e.g., Chawla et al., 1992], antibacterial [Sridhar et al., 2005], antidiarrheal [Agunu et al., 2005], antihistamine [Alam et al., 2002], analgesic [Dharmasiri et al., 2003], antiviral [Goncalves et al., 2001], antioxidant [Gallo et al., 2006], antitrypanosomal [Kiuchi et al., 2004], antifungal [Hernandez et al., 1999], anticonvulsant [Tandon & Gupta, 2005], cytotoxic [Hirobe et al., 1997], antiandrogenic [Das et al., 2004], insecticidal [Pushpalatha & Muthukrishnan, 1995], antivenin [Alam & Gomes, 2003], and profertility agents [Eckman & Hines, 1993]. The most widely cited effects are those of V. agnus-castus (VAC), the fruits of which are cited in both historical and modern usage for their impact on the reproductive physiology of one or both sexes [Daniele et al., 2005]. Fruit extracts of VAC induce increases in female progesterone levels and are widely employed as natural remedies for premenstrual syndrome and menstrual irregularities [Girman et al., 2002; Tesch, 2001; Wuttke et al., 2003]. The German government has approved its use for treatment of premenstrual syndrome, breast pain, and irregular menstrual cycles [Hardy, 2000]. Double-blind studies in that country found that Vitex extract was effective in treatment of luteal deficits [Gerhard et al., 1998; Milewicz et al., 1993] and premenstrual syndrome [Schellenberg, 2001], presumably as a result of its effects on progesterone levels. VAC's effects on male reproductive physiology are suggested by its traditional use among monks to reduce libido (hence the common name "chasteberry"), but clinical data are lacking.

Chimpanzees in the Gombe National Park in Tanzania naturally consume the fruits of a related species, V. fischeri, during its short annual fruiting season [Goodall, 1968]. The phytochemistry of this Vitex species has not yet been described. Here, we test for potential effects of V. fischeri consumption on reproductive hormones in both male and female chimpanzees at Gombe.

METHODS

Study Location Community

The Kasekela community of chimpanzees in Gombe National Park, Tanzania, has been the subject of continuous research since 1960 [Goodall, 1986]. The park, spanning 35 km^2 along the shores of Lake Tanganyika, is composed of a mosaic of grassland and semideciduous and evergreen forests extending from elevations of 770–1,600 m [Goodall, 1986; Wrangham, 1977; Pussey et al., 2008]. In the beginning of 2002, the Kasekela community contained 51 individuals, including 20 adult and subadult females and 12 adult and subadult males. Research with Gombe chimpanzees was conducted in compliance with the Institutional Animal Care and Use Committees at Harvard University and the University of Minnesota and with permission from Tanzanian governmental authorities.

Behavioral Sampling

Field researchers at Gombe conduct daily nestto-nest observations of Kasekela chimpanzees. If possible, one focal individual is followed throughout the day; feeding and social behavior of the focal are recorded continuously and party membership is recorded every 15 min. The plant part and species of all items eaten are recorded. A Tanzanian botanist (G. G.) maintains an on-site herbarium collection and helps with identification of plant species. We calculated dietary composition based on the items consumed by focal animal at the time of each 15-min scan sample. Note that the dates of the Vitex season were defined by the first and last observations of any focal animal consuming the fruit and that observers traveled with focal animals rather than observing Vitex trees full time. Our field data do not permit us to calculate the dates and intensity of Vitex feeding by individual chimpanzees.

Endocrine Sampling and Laboratory Analysis

Between June 2001 and December 2002, we collected fresh, uncontaminated samples by placing a plastic sheet underneath a known subject or by pipetting from vegetation. Samples were frozen as quickly as possible, within a time frame (0-14 hr) that has been demonstrated not to significantly affect ovarian hormone or creatinine concentrations [Kesner et al., 1995].

M. E. T. assayed urine samples in the Primate Reproductive Ecology Laboratory at Harvard University using competitive enzyme-immunoassay (reagents c/o, C. J. Munro, University of California, Davis). Female urine samples were assayed for the urinary metabolites of estrogen (estrone conjugates, E_1C) and progesterone (pregnanediol-3-glucuronide, PdG). We assayed male samples for testosterone after conducting a hydrolysis and extraction procedure to deconjugate the key urinary metabolite, testosterone glucuronide [Mohle et al., 2002; Muller & Wrangham, 2004]. Enzyme-immunoassay procedures followed previously published protocols [e.g., Czekala et al., 1986; Emery Thompson, 2005a; Shideler et al., 1990] with coating dilutions for Nunc Maxisorp immunoplates at 1:6,000 for estrone- β -glucuronide (Antibody

R522-2), 1:30,000 for PdG (Pab 13904), and 1:10,000 for test osterone (R156-7). We standardized all results for creatinine (Cr) concentration to adjust for sample concentration [Taussky, 1954]. Interassay coefficients of variation (CVs) were 13.6% for E₁C (n = 56), 14.2% for PdG (n = 26), and 11.5% for test osterone (n = 17). Intraassay CVs from 6 replicate urine samples were 4.9% for E₁C, 3.4% for PdG, and 5.6% for test osterone.

Data Analysis

For males, we considered morning (before 10 am) and "afternoon" (after 10 am) samples separately owing to circadian effects on urinary testosterone concentrations in chimpanzees [Muller & Lipson, 2003]. We included both adult and subadult (>10)years of age) males. For females, we categorized each urine sample according to the reproductive state of the subject based on birth records, sexual swelling occurrence, and pregnancy testing. In order to obtain a valid statistical comparison of hormonal levels in females of the same reproductive status before, during, and after Vitex season, we limited our analysis primarily to noncycling females. This included adolescent females at least 8 years of age and females in the period of lactational amenorrhea before resumption of cycling. To test for significant changes in hormonal values, we used Wilcoxon matched-pairs tests ($\alpha = 0.05$, two-tailed significance) to compare individual chimpanzees' average values over the 6month period before the first observation of Vitex consumption with their average values during Vitex consumption (first to last observation: 17 February-26 March 2002). We also compared paired values from the period of *Vitex* consumption and average levels in the 6 months after the last *Vitex* feeding observation.

RESULTS

Kasekela chimpanzees were observed consuming V. fischeri fruits (Fig. 1) between 17 February 2002 and 26 March 2002. During this time period, 1,547 scan observations were recorded (approximately 387 observation hours), with focal animals feeding for 797 (51.5%) of these observation points. Fruits of V. fischeri were the most common dietary item for the chimpanzees over this period, accounting for 17.3% of feeding observations. Seventeen of 25 known Kasekela females and 10 of 12 known males were observed in feeding parties at V. fischeri trees.

We compared mean urinary reproductive hormone concentrations in male and female chimpanzees during the *Vitex* season with mean concentrations from the same individuals during the 6 months preceding and the 6 months following the season. Urinary progestin levels of Gombe females were significantly elevated in the *Vitex* season compared with the previous 6-month period (Wilcoxon signedranks test, z = -2.023, N = 5 females, P = 0.043) and the subsequent 6-month period (z = -2.521, N = 8,

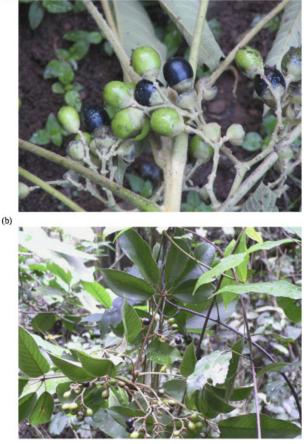


Fig. 1. Photos of *Vitex fischeri* from Gombe National Park, illustrating (\mathbf{a}) ripe (black) and unripe (green) fruits and (\mathbf{b}) mature leaves with fruits (photos: Michael Wilson).

P = 0.012, Fig. 2a). Elevated values were tightly tied to the dates of our Vitex feeding observations and fell outside of the typical range of variation for PdG (Fig. 3). PdG levels of noncycling females during the Vitex season averaged $4,130\pm923$ ng/mg-Cr (mean \pm standard error, N = 5 females), more than four standard deviations above their typical levels $(810 \pm 232 \text{ ng/mg-})$ Cr, N = 14 females sampled from June 2001 to December 2002). We were only able to collect seven samples from four cycling females during the Vitex season; all were collected during the presumed follicular phase [i.e., during maximal swelling or 10 days earlier; Emery Thompson, 2005a], a time of characteristically low progesterone activity. These samples averaged $9,121 \pm 4,519$ ng/mg-Cr, three standard deviations above their usual levels $(1,964 \pm$ 765 ng/mg-Cr, N = 10 females).

In contrast, urinary estrogen levels of females during the *Vitex* season were not significantly different from values during the previous 6 months (z = -1.214, N = 5, P = 0.225), which were slightly higher on average, and values during the following 6 months (z = -1.680, N = 8, P = 0.093), which were slightly lower (Fig. 2b).

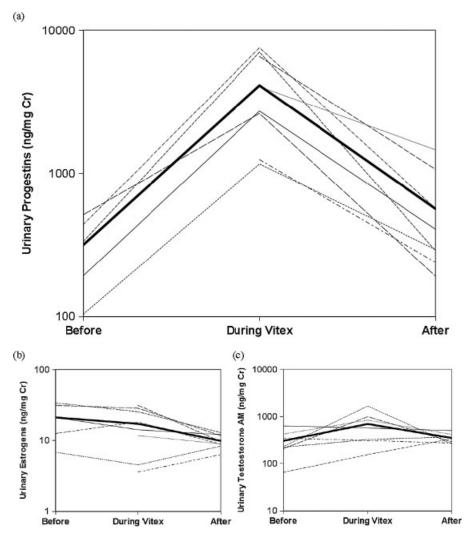


Fig. 2. Urinary hormone measurements of chimpanzees during the *Vitex* season (17 February 02–26 March 02) compared with the 6 months before and 6 months after. Each thin solid or dotted line indicates an individual chimpanzee, whereas the solid thicker line denotes the mean across individuals. Y-axes use a log-scale to accommodate large effects. (a) Urinary PdG levels of noncycling females; (b) urinary E_1C levels of noncycling females; and (c) urinary testosterone of males in morning samples. PdG, pregnanediol-3-glucuronide; E_1C , estrone conjugates.

Male urinary testosterone levels were also not significantly affected by *Vitex* feeding. Neither morning nor afternoon testosterone levels differed significantly from the previous 6 months (morning: z = -1.859, N = 7 males, P = 0.063; afternoon: z = -1.352, N = 6, P = 0.176) or the following 6 months (morning: z = -0.944, N = 7, P = 0.345; afternoon: z = -0.280, N = 9, P = 0.779, Fig. 2b). Testosterone levels were slightly elevated on average, which contrasts with the prediction based on VAC's purported impact on male libido.

DISCUSSION

The consumption of *V. fischeri* fruits coincided with an abrupt and dramatic elevation in progesterone levels of female chimpanzees at Gombe. This effect parallels the increase in progesterone observed from the consumption of VAC extracts by human females. Precise causes of these effects are not known, but at least in the case of VAC do not appear to be related directly to a progesterone-like compound in the plant. Extracts of VAC fruits increase progesterone production in women via upregulation of progesterone receptors and modulation of the folliclestimulating hormone/luteinizing hormone ratio [Girman et al., 2002; Liu et al., 2001; Tesch, 2001]. Reproductive hormones may also be altered through VAC's dopaminergic and antiprolactinemic effects [Meier et al., 2000; Sliutz et al., 1993; Tesch, 2001; Wuttke et al., 2003] and via competitive binding to estrogen α and β receptors [Liu et al., 2001, 2004].

The described effects of VAC consumption are primarily beneficial for fecundity in women [Girman et al., 2002; Milewicz et al., 1993; Schellenberg, 2001; Tesch, 2001; Wuttke et al., 2003]. This contrasts with

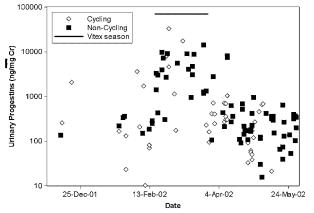


Fig. 3. Scatterplot of the urinary PdG concentrations of individual samples collected from cycling (open diamond) and noncycling (closed square) females, illustrating the close relationship between *Vitex fischeri* consumption and dramatically increased progestin levels. The period of *V. fischeri* feeding observations is noted by a solid black line. PdG, pregnanediol-3-glucuronide.

the hypothesized effects of many phytoestrogen compounds in plants, as well as the observed negative effects of such compounds on reproduction in herbivores and other species [Adams, 1995; Hughes, 1988; Leopold et al., 1976; Trisomboon et al., 2004]. Although many phytoestrogens appear to have positive effects on health, particularly on prevention of cancers and cardiovascular disease [Ososki & Kennelly, 2003], their effects on reproductive health and fertility are frequently negative or inhibitory [Jefferson & Newbold, 2000; Whitten & Patisaul, 2001]. In our study, estrogen levels of chimpanzees were not significantly outside of the normal range of variation during V. fischeri consumption. We do not know whether the observed effects on progesterone levels in chimpanzees affected fecundity, positively or negatively. The occurrence of sexual swellings was relatively high but not unusual, and no conceptions occurred during the *Vitex* season or during the following 6 months. Given long interbirth intervals and the typical seasonal variation in chimpanzee reproductive function [Anderson et al., 2006; Emery Thompson, 2005a; Emery Thompson & Wrangham, 2008; Wallis, 1995, 2002], the potential effect of Vitex consumption on fecundity is difficult to evaluate. However, Vitex has been used annually by Gombe chimpanzees since at least 1974 and the feeding season is often longer or more intense than it was in 2002 [Gombe Stream Research Centre, unpublished data]. Such habitual use would seem to be disfavored were the fruit to produce strong negative fertility effects. However, it is also possible that the duration of altered hormonal function during any given Vitex season is insufficient to produce significant disruptions or enhancements in female fertility.

Reproductive hormones may by influenced by the diet through other pathways. In chimpanzees, as

in humans [Ellison et al., 1993] and orangutans [Knott, 2001], ovarian hormone levels correlate with increased consumption of foods high in nutritional quality [Emery Thompson & Wrangham, 2008; Emery Thompson et al., 2007]. However, this is unlikely to be the cause of the hormonal effects we found because overall ripe fruit consumption during February and March 2002 was relatively low. Furthermore, in chimpanzees (including those at Gombe), the acute effects of increased dietary quality are seen primarily in increasing estrogen levels, whereas progesterone responses are often dampened or delayed [Emery Thompson, 2005b; Emery Thompson & Wrangham, 2008]. The responses observed here also far exceed those observed during seasonal shifts in food availability for chimpanzees.

VAC consumption has historically been linked with reduced male sexual motivation and has been used by monks for this purpose [Daniele et al., 2005]. However, we are not aware of any clinical evidence to suggest effects on testicular function in humans. In our study of *V. fischeri* consumption, we did not detect significant changes in the testosterone levels of male chimpanzees.

Vitex species feature prominently in the diets of chimpanzees [V. sp.: Azuma & Toyoshima, 1961; V. doniana: Duvall, 2000; Moscovice et al., 2007; Sugiyama & Koman, 1987; Tutin & Fernandez, 1993; V. madiensis: McGrew et al., 1988; V. grandifolia: Poulsen et al., 2001; V. cienkowski: Sugiyama & Koman, 1987; V. ferruginea: Suzuki, 1969], bonobos [V. sp.: Badrian et al., 1981], gorillas [V. sp.: Cousins & Huffman, 2002; Rogers et al., 2004; V. doniana: Nishihara, 1995; Rogers et al., 1990; Williamson et al., 1990; V. grandifolia: Poulsen et al., 2001], and other primates [V. grandifolia: Poulsen et al., 2001] in sites across Africa. Like V. fischeri, the chemical compositions of these Vitex species are as yet undescribed. There is therefore ample potential to study whether phytochemical influences of Vitex on female hormone levels are a more general phenomenon, and whether broader reproductive effects result from it, particularly where usage is more intense. Such further evidence would raise the intriguing possibility that primates select these plants for their positive phytochemical rather than nutritional properties, just as they consume plants with particular mechanical or chemical properties for apparent medicinal purposes [Carrai et al., 2003; Huffman, 1997].

ACKNOWLEDGMENTS

We thank Dr. Shadrack Kamenya, Dr. D. Anthony Collins, and the field staff of Gombe Stream Research Centre, who maintained daily data collections. We are grateful to the Jane Goodall Institute for supporting long-term field research. Archiving, database development, and analysis of the long-term data were supported by the University of Minnesota, the US National Science Foundation, the Jane Goodall Institute, and the Harris Steel Group. Laboratory analyses were supported by grants to MET from the Harvard University, the Wenner-Gren Foundation, and the L. S. B. Leakey Foundation. Research at Gombe National Park was conducted with permission from the Tanzanian Council for Science and Technology, Tanzanian National Parks, and the Tanzanian Wildlife Research Institute. All research was conducted in compliance with Institutional Animal Care and Use Committee regulations at the authors' home institutions.

REFERENCES

- Adams N. 1995. Detection of the effects of phytoestrogens on sheep and cattle. J Anim Sci 73:1509–1515.
- Agunu A, Yusuf S, Andrew GO, Zezi A, Adbdurahman EM. 2005. Evaluation of five medicinal plants used in diarrhoea treatment in Nigeria. J Ethnopharmacol 101:27–30.
- Alam M, Gomes A. 2003. Snake venom neutralization by Indian medicinal plants (*Vitex negundo* and *Emblica* officinalis) root extracts. J Ethnopharmacol 86:75–80.
- Alam G, Wahyuono S, Ganjar IG, Hakim L, Timmerman H, Verpoorte R. 2002. Tracheospasmolytic activity of viteosin-A and vitexicarpin isolated from *Vitex trifolia*. Planta Med 68:1047-1049.
- Anderson DP, Nordheim EV, Boesch C. 2006. Environmental factors influencing the seasonality of estrus in chimpanzees. Primates 47:43–50.
- Azuma S, Toyoshima A. 1961. Progress report of the survey of chimpanzees in their natural habitat, Kabogo Point area, Tanganyika. Primates 3:61–70.
- Badrian N, Badrian A, Susman RL. 1981. Preliminary observations on the feeding behavior of *Pan paniscus* in the Lomako Forest of central Zaire. Primates 22:173–181.
- Bhargava S. 1989. Antiandrogenic effects of a flavonoid-rich fraction of *Vitex negundo* seeds: a histological and biochemical study in dogs. J Ethnopharmacol 27:327–339.
- Carrai V, Borgognini-Tarli SM, Huffman MA, Bardi M. 2003. Increase in tannin consumption by sifaka (*Propithecus verreauxi verreauxi*) during the birth season: a case for self-medication in primates? Primates 44:61–66.
- Chawla A, Sharma A, Handa S, Dhar K. 1992. Chemical investigation and anti-inflammatory activity of *Vitex negun*do seeds. J Nat Prod 55:163–167.
- Cousins D, Huffman MA. 2002. Medicinal properties in the diet of gorillas: an ethno-pharmacological evaluation. Afr Study Monogr 23:65–89.
- Czekala NM, Gallusser S, Meier J, Lasley BL. 1986. The development and application of an enzyme immunoassay for urinary estrone conjugates. Zoo Biol 5:1–6.
- Daniele C, Coon JT, Pittler MH, Ernst E. 2005. Vitex agnuscastus: a systematic review of adverse events. Drug Saf 28:319–332.
- Das S, Parveen S, Kundra CP, Pereira BM. 2004. Reproduction in male rats is vulnerable to treatment with the flavonoid-rich seed extracts of *Vitex negundo*. Phytother Res 18:8–13.
- Dharmasiri M, Jayakody J, Galhena G, Liyanage S, Ratnasooriya W. 2003. Anti-inflammatory and analgesic activities of mature fresh leaves of *Vitex negundo*. J Ethnopharmacol 87:199–206.
- Duvall CS. 2000. Important habitat for chimpanzees in Mali. Afr Study Monogr 21:173–203.

- Eckman K, Hines DA. 1993. Indigenous multipurpose trees of Tanzania: uses and economic benefits for people. Food and Agriculture Department of the United Nations.
- Ellison PT, Panter-Brick C, Lipson SF, O'Rourke MT. 1993. The ecological context of human ovarian function. Hum Reprod 8:2248–2258.
- Emery Thompson M. 2005a. Reproductive endocrinology of wild female chimpanzees (*Pan troglodytes schweinfurthii*): methodological considerations and the role of hormones in sex and conception. Am J Primatol 67:137–158.
- Emery Thompson M. 2005b. Endocrinology and ecology of wild female chimpanzee reproduction. PhD dissertation, Harvard University, Cambridge, MA.
- Emery Thompson M, Wrangham RW. 2008. Diet and reproductive function in wild female chimpanzees (*Pan troglodytes schweinfurthii*) at Kibale National Park, Uganda. Am J Phys Anthropol 135:171–181.
- Emery Thompson M, Kahlenberg SM, Gilby IC, Wrangham RW. 2007. Core area quality is associated with variance in reproductive success among female chimpanzees at Kanyawara, Kibale National Park. Anim Behav 73:501–512.
- Foth D, Cline J. 1998. Effects of mammalian and plant estrogens on mammary glands and uteri of macaques. Am J Clin Nutr 68:1413S-1417S.
- Gallo MBC, Beltrame FL, Vieira PC, Cass QB, Fernandes JB, da Silva MFDGF. 2006. Quantitative determination of 20hydroxyecdysone in methanolic extract of twigs from *Vitex polygama* Cham. J Chromatogr B Biomed Sci Appl 832:36–40.
- Garey J, Markiewicz L, Gurpide E. 1992. Estrogenic flowers, a stimulus for mating activity in female vervet monkeys. XIVth Congress of the International Primatological Society Abstracts, Strassbourg, p 210.
- Gerhard I, Patek A, Monga B, Blank A, Gorkow C. 1998. Mastodynon for female infertility: randomized, placebocontrolled, clinical double-blind study. Res Complement Med 5:272-278.
- Girman A, Lee R, Klingler B. 2002. An integrative medicine approach to premenstrual syndrome. Am J Obstet Gynecol 188:S56–S65.
- Goncalves J, Leitao S, Monarche F, Miranda M, Santos M, Romanos M, Wigg M. 2001. In vitro effect of flavonoid-rich extracts of *Vitex polygama* (Verbenaceae) against acyclovirresistant herpes simplex virus type 1. Phytomedicine 8:477-480.
- Goodall J. 1968. Behaviour of free-living chimpanzees of the Gombe Stream area. Anim Behav Monogr 1:163–311.
- Goodall J. 1986. The chimpanzees of Gombe: patterns of behavior. Cambridge: Belknap Press.
- Hardy M. 2000. Women's health series: herbs of special interest to women. J Am Pharm Assoc 40:234-242.
- Harrison RM, Phillippi PP, Swan KF, Henson MC. 1999. Effect of genistein on steroid hormone production in the pregnant rhesus monkey. Proc Soc Exp Biol Med 222:78–84.
- Hernandez M, Heraso C, Villarreal M, Vargas-Arispuro I, Aranda E. 1999. Biological activities of crude plant extracts from *Vitex trifolia* L. (Verbenaceae). J Ethnopharmacol 67:37-44.
- Higham JP, Ross C, Warren Y, Heistermann M, MacLaron AM. 2007. Reduced reproductive function in wild baboons (*Papio hamadryas anubis*) related to natural consumption of the African black plum (*Vitex doniana*). Horm Beh 52:384–390.
- Hirobe C, Qiao Z, Takeya K, Itokawa H. 1997. Cytotoxic flavonoids from *Vitex agnus-castus*. Phytochemistry 46:521–524.
- Hopkins W, Bailey J, Fuller G. 1977. Hormone effects of zearalenone in nonhuman primates. J Toxicol Environ Health 3:43–57.
- Huffman MA. 1997. Current evidence for self-medication in primates: an interdisciplinary perspective. Yrbk Phys Anthropol 40:171–200.

- Hughes CL. 1988. Phytochemical mimicry of reproductive hormones and modulation of herbivore fertility by phytoestrogens. Environ Health Perspect 78:171–174.
- Jefferson WN, Newbold RR. 2000. Potential endocrinemodulating effects of various phytoestrogens in the diet. Nutrition 16:658–662.
- Kesner JS, Knecht EA, Krieg Jr EF. 1995. Stability of urinary female reproductive hormones stored under various conditions. Reprod Toxicol 9:239–244.
- Kiuchi F, Matsuo K, Ito M, Qui TK, Honda G. 2004. New norditerpenoids with trypanocidal activity from *Vitex trifolia*. Chem Pharm Bull (Tokyo) 52:1492–1494.
- Knott CD. 2001. Ape models of female reproductive ecology. In: Ellison PT, editor. Reproductive ecology and human evolution. Chicago: Aldine. p 429–463.
- Leopold A, Erwin M, Oh J, Browning B. 1976. Phytoestrogens: adverse effects on reproduction in California quail. Science 191:98–100.
- Li W, Cui C, Cai B, Wang H, Yao Z. 2005. Flavonoids from *Vitex trifolia* L. inhibit cell cycle progression at G2/M phase and induce apoptosis in mammalian cancer cells. J Asian Nat Prod Res 7:615–626.
- Liu J, Burdette J, Zu H, Gu C, van Breemen R, Bhat K, Booth N, Constantinou A, Pezzuto J, Fong H, Farnsworth N, Bolton J. 2001. Evaluation of estrogenic activity of plant extracts for the potential treatment of menopausal symptoms. J Agric Food Chem 49:2472–2479.
- Liu J, Burdette J, Sun Y, Deng S, Schlecht S, Zheng W, Nikolic D, Mahady G, van Breemen R, Fong H, Pezzuto J, Bolton J, Farnsworth N. 2004. Isolation of linoleic acid as an estrogenic compound from the fruits of *Vitex agnus-castus* L. (chaste-berry). Phytomedicine 11:18–23.
- McGrew WC, Baldwin P, Tutin CE. 1988. Diet of wild chimpanzees (*Pan troglodytes verus*) at Mt. Assirik, Senegal: I. Composition. Am J Primatol 16:213–226.
- Meier B, Berger D, Hoberg E, Sticher O, Schaffner W. 2000. Pharmacological activities of *Vitex agnus-castus* extracts in vitro. Phytomedicine 7:373–381.
- Milewicz A, Gejdel E, Sworen H, Sienkiewicz K, Jedrzejak J, Teucher T, Schmitz H. 1993. *Vitex agnus castus* extract in the treatment of luteal phase defects due to latent hyperprolactinemia. Results of a randomized placebo-controlled double-blind study. Arzneimittelforschung 43: 752–756.
- Mohle U, Heistermann M, Palme R, Hodges J. 2002. Characterization of urinary and fecal metabolites of testosterone and their measurement for assessing gonadal endocrine function in male nonhuman primates. Gen Comp Endocrinol 129:135–145.
- Moscovice L, Issa M, Petrzelkova K, Keuler N, Snowdon CT, Huffman MA. 2007. Fruit availability, chimpanzee diet, and grouping patterns on Rubondo Island, Tanzania. Am J Primatol 69:487–502.
- Muller MN, Lipson SF. 2003. Diurnal patterns of urinary steroid excretion in wild chimpanzees. Am J Primatol 60:161–166.
- Muller MN, Wrangham RW. 2004. Dominance, aggression and testosterone in wild chimpanzees: a test of the 'challenge hypothesis'. Anim Behav 67:113–123.
- Nishihara T. 1995. Feeding ecology of western lowland gorillas in the Nouabale-Ndoki National Park, Congo. Primates 36:151–168.
- Okuyama E, Fujimori S, Yamazaki M, Deyama T. 1998. Pharmacologically active components of viticis fructus (*Vitex rotundifolia*). II. The components having analgesic effects. Chem Pharm Bull (Tokyo) 46:655–662.
- Ososki AL, Kennelly EJ. 2003. Phytoestrogens: a review of the present state of research. Phytother Res 17:845–869.
- Poulsen JR, Clark CJ, Smith TB. 2001. Seed dispersal by a diurnal primate community in the Dja Reserve, Cameroon. J Trop Ecol 17:787–808.

- Pusey AE, Wilson ML, Collins DA. 2008. Human impacts, disease risk, and population dynamics in the chimpanzees of Gombe National Park, Tanzania. Am J Primatol 70:738–744.
- Pushpalatha E, Muthukrishnan J. 1995. Larvicidal activity of a few plant extracts against *Culex quinquefasciatus* and *Anopheles stephensi*. Indian J Malariol 32:14–23.
- Rogers M, Maisels F, Williamson E, Fernandez M, Tutin CE. 1990. Gorilla diet in the Lope Reserve, Gabon: a nutritional analysis. Oecologia 84:326–339.
- Rogers ME, Abernethy KA, Bermejo M, Cipolletta C, Doran D, McFarland K, Nishihara T, Remis M, Tutin CE. 2004. Western gorilla diet: a synthesis from six sites. Am J Primatol 64:173–192.
- Schellenberg R. 2001. Treatment for the premenstrual syndrome with agnus castus fruit extract: prospective, randomised, placebo controlled study. Br J Med 322: 134–137.
- Shideler SE, Munro CJ, Tell L, Owitt G, Laughlin L, Chatterton Jr R, Lasley BL. 1990. The relationship of serum estradiol and progesterone concentrations to the enzyme immunoassay measurements of urinary estrone conjugates and immunoreactive pregnanediol-3-glucuronide in *Macaca mulatta*. Am J Primatol 22:113–122.
- Sliutz G, Spieser P, Schultz A, Spona J, Zeillinger R. 1993. Agnus castus extracts inhibit prolactin secretion of rat pituitary cells. Horm Metab Res 25:253–255.
- Sridhar C, Rao KV, Subbaraju G. 2005. Flavonoids, triterpenoids and a lignan from *Vitex altissima*. Phytochemistry 66:1707–1712.
- Strier KB. 1993. Menu for a monkey. Nat Hist 102:34-43.
- Sugiyama Y, Koman J. 1987. A preliminary list of chimpanzees' alimentation at Bossou, Guinea. Primates 28:133–147.
- Suksamrarn A, Kumpun S, Kirtikara K, Yingyongnarongkul B, Kuksamrarn S. 2002. Iridoids with anti-inflammatory activity from *Vitex penuncularis*. Planta Med 68:72–73.
- Suzuki A. 1969. An ecological study of chimpanzees in a savanna woodland. Primates 10:103–148.
- Tandon V, Gupta R. 2005. An experimental evaluation of antoconvulsant activity of *Vitex negundo*. Indian J Physiol Pharmacol 49:199–205.
- Taussky H. 1954. A microcolorimetric determination of creatinine in urine by the Jaffe reaction. J Biol Chem 208: 853–861.
- Tesch BJ. 2001. Herbs commonly used by women: an evidencebased review. Am J Obstet Gynecol 188:S44–S55.
- Trisomboon H, Malaivijitnond S, Watanabe G, Taya K. 2004. Estrogenic effects of *Pueraria mirifica* on the menstrual cycle and hormone-related ovarian functions in cycling female cynomolgus monkeys. J Pharmacol Sci 94:51–59.
- Tutin CEG, Fernandez M. 1993. Composition of the diet of chimpanzees and comparisons with that of sympatric lowland gorillas in the Lope Reserve, Gabon. Am J Primatol 30:195–211.
- Wallis J. 1995. Seasonal influence on reproduction in chimpanzees of Gombe National Park. Int J Primatol 16: 435–451.
- Wallis J. 1997. A survey of reproductive parameters in the free-ranging chimpanzees of Gombe National Park. J Reprod Fertil 109:297–307.
- Wallis J. 2002. Seasonal aspects of reproduction and sexual behavior in two chimpanzee populations: a comparison of Gombe (Tanzania) and Budongo (Uganda). In: Boesch C, Hohmann G, Marchant LF, editors. Behavioural diversity in chimpanzees and bonobos. Cambridge, MA: Cambridge University Press. p 181–191.
- Whitten P. 1983. Flowers, fertility, and females. Am J Phys Anth 60:269–270.
- Whitten PL, Naftolin F. 1998. Reproductive actions of phytoestrogens. Baillieres Clin Endocrinol Metab 12: 667–690.

- Whitten PL, Patisaul HB. 2001. Cross-species and interassay comparisons of phytoestrogen action. Environ Health Perspect Suppl 109:5–20.
- Williamson E, Tutin CEG, Rogers ME, Fernandez M. 1990. Composition of the diet of lowland gorillas at Lope in Gabon. Am J Primatol 21:265–277.
- Wood C, Kaplan J, Stute P, Cline J. 2006. Effects of soy on the mammary glands of premenopausal female monkeys. Fertil Steril 85:1179–1186.
- Wrangham RW. 1977. Feeding behaviour of chimpanzees in Gombe National Park, Tanzania. In: Clutton-Brock TH, editor. Primate ecology: studies of feeding and ranging behaviour in lemurs, monkeys and apes. New York: Academic Press. p 503–537.
- Academic Press. p 503–537.
 Wuttke W, Jarry H, Christoffel V, Spengler B, Seidlova-Wuttke D. 2003. Chaste tree (*Vitex agnus-castus*)—pharmacology and clinical indications. Phytomedicine 10:348–357.